

## Effects of GA<sub>3</sub> Concentrations and Mechanical Scarification on Germination and Seedling Growth Performance of Sour Orange (*Citrus aurantium* L.) Rootstock

Raghib Mohammad Ghani<sup>1\*</sup>, Sadat Mohammad Ismail<sup>1</sup>, Zabihullah Safi<sup>1</sup>, Hameedi Atal<sup>2</sup>, and Sediqli Naveedullah<sup>3</sup>

<sup>1</sup>Department of Horticulture, Faculty of Agriculture, Nangarhar University, Nangarhar, Afghanistan

<sup>2</sup>Department of Horticulture, Faculty of Agriculture, Sayed Jammaluddin Afghani University, Kunar, Afghanistan

<sup>3</sup>Department of Horticulture, Faculty of Agriculture, Alberoni University, Kapisa, Afghanistan

\*Corresponding author: [ghaniraghib@gmail.com](mailto:ghaniraghib@gmail.com)

### ABSTRACT

The current experiment was conducted in Agriculture Faculty Lab (pro trays), at Nangarhar University, Afghanistan. As the germination of the sour orange seed is very poor and needs a long time for their competent germination, it was aimed to enhance germination and produce standard seedlings for proper growth and development. The experiment was laid out in a completely randomized design (CRD) having six treatments, T<sub>1</sub> (50 ppm + scarification), T<sub>2</sub> (100 ppm + scarification), T<sub>3</sub> (150 ppm + scarification), T<sub>4</sub> (200 ppm + scarification), T<sub>5</sub> (250 ppm + scarification) and T<sub>6</sub> (Control) with three replications. The scarification was done manually for seed coat removal after the extraction of seeds from fruits. Analysis of variance revealed to be significant for most of the characters under study. The study revealed that among all treatments, T<sub>5</sub> (250 ppm + scarification) was the best treatment for growth and root parameters, followed by T<sub>4</sub> (200 ppm + scarification) while the lowest of growth and root parameters were registered in T<sub>6</sub> (control). This investigation confirmed that high concentration of GA<sub>3</sub> significantly affected the seed germination of sour orange.

**Keywords:** Gibberellic Acid, Mechanical Scarification, Sour Orange

### INTRODUCTION

Citrus is a genus of flowering trees having numerous species belonging to the family Rutaceae with chromosome number ( $2n = 2x = 18$ ) experiencing more diversity in the species. Citrus is one of the most extensively cultivated fruit crops in the world and a substantial part of the human diet among fruit crops (Liu et al., 2012). Cultivated species of citrus fruit crops are the large group, containing approximately 16 to 156 species around the world, and have spread throughout the globe (Ollitrault & Navaro, 2012). In addition, citrus have relative complexity in their broad morphological characters, which caused difficulty in their identification. Sour orange (*Citrus aurantium* L.) is considered one of the most popular rootstocks for different citrus species especially for sweet oranges in Eastern Zone of Afghanistan. This rootstock can significantly influence scion growth and development, fruit size, fruit skin, fruit quality, fruit acidity, and resistance to biotic and abiotic stresses. Rootstock is one of the important and essential components of budded /grafted fruit plants, which determine the success, or failure of the established orchards, apart from the influence on the plant vigour, size, and longevity of the tree, it helps in the early production of quality fruits (Yadav et al., 2022).

To make the scion adapted to wide range of biotic and abiotic conditions; selection of the competent rootstock is the most crucial method to overcome the adverse effects of climate, soil, and resistance to certain diseases and pests. However, potent struggles have always been made to increase environmental adaptability,

resistance to biotic stress (Gummosis / *Phytophthora* foot rot, quick decline or Citrus tristeza) and abiotic stress (Drought, Cold, Heat, Salinity and Light), enhanced nutrient uptake by using the appropriate scion and rootstock combinations in citrus fruit crops propagation (Sharaf *et al.*, 2016).

The plants of this rootstock are medium in size, early maturing, and prolific with quality produce and make a good union with sweet oranges. It is widely used across the globe and show resistance to gummosis caused by *Phytophthora*, while susceptible against Citrus Tristeza Virus (CTV).

Citrus seeds of different rootstocks are observed to have poor germination, poor seedling uniformity, and uneven growth performance and take a long time for budding/grafting stages. Thus, it is highly required to enhance germination, uniformity, and rapid establishment for budding or grafting, by choosing suitable rootstock (Castle, 1982). GA<sub>3</sub> is considered an important germination promoter, which increase, seed germination and proper root development (Zanotti & Barros, 2014). Gibberellin hormone is one of the most popular phytohormones that plays an important role in multifunction, initial plant development, and promote germination. In recent years, gibberellic acid has been used widely as a synthetic hormone or plant growth regulator to promote and stimulate seed germination and seedling stem elongation (Khatana, 2010; Stejskalová & Kupka, 2015). Gibberellin enhances seed germination by the mean of Alpha- amylase enzyme break down to the starch and glucose into smaller sugar contents to promote embryo growth until it becomes autotrophic (Vieira, 2002). Gibberellic acid encourages the growth of seeds that increases the growth rates in sweet orange (Hoda, 2010). This rootstock is commonly obtained through seed but seed germination and proper development is not up to the mark. Keeping these points in mind, an investigation has been conducted to study the effects of GA<sub>3</sub> concentrations and mechanical scarification on germination and seedling growth performance of sour orange (*Citrus aurantium* L.) rootstock.

## MATERIALS AND METHODS

This experiment was laid out in February 2022 in the Agriculture Faculty Lab (pro trays), Nangarhar University; Afghanistan. Geographically, the research farm is situated between 34° 28' 38.40" North latitude and 70° 22' 0.141" East longitude. It is situated at about 600 meters above the mean sea level. The full-ripened and colored fruits were brought from the sour orange orchards of the University campus, then disease-free, uniform-sized fruits were selected, and seeds were extracted from fruits and cleaned with the help of tap water. After extraction, seeds were spread out on cotton cloth to dry and remove excessive water and moisture. The study was conducted in a completely randomized design having six treatments T<sub>1</sub> (50 ppm + scarification), T<sub>2</sub> (100 ppm + scarification), T<sub>3</sub> (150 ppm + scarification), T<sub>4</sub> (200 ppm + scarification), T<sub>5</sub> (250 ppm + scarification) and T<sub>6</sub> (Control = distilled water + no scarification), with three replications. Gibberellic acid (Duchefa biochemical, assay > 90% ) was weighed (50, 100,150,200, and 250) mg separately by helping electronic balance (jewelry scale, model 1020), then dissolved every required amount of GA<sub>3</sub> powder separately in 10 ml of 99% pure ethyl alcohol in different beakers and distilled water was added to make up the volume equal to (1000) ml or 1 liter to obtain desired concentrations of (50, 100, 150, 200 and 250 ppm GA<sub>3</sub> solution and then seeds were kept for 10 minutes (Quick Deeping method) in every solution.

All prepared seeds were subjected to mechanical scarification and 100 seeds were used for each treatment and then were kept in GA<sub>3</sub> solutions for 10 minutes. The final count of germination percentage was recorded on 20<sup>th</sup> day of germination test. Five competitive plants were chosen randomly and tagged to keep a record of other observations. To determine the mean values of the parameters, the data collected for various plant properties

were averaged out independently. Data related to different parameters e.i. initial emergences, germination (%), seedling height (cm), stem or diameter (mm), length of root (cm), and number of roots/plant, were recorded.

The data of the present experiment was subjected to statistical analysis in completely randomized Design (CRD). With one-way, ANOVA and then the treatment means were compared by using Critical Differences test at  $P \leq 0.05$ .

## RESULTS

### Germination percentage and Initial emergence

The attentive study of the results depicted in (Table 1) divulged that the combined effect of GA<sub>3</sub> and mechanical scarification enhanced days taken for initial germination, and germination percentage. The highest germination percentage (96.67 %) was recorded in T<sub>5</sub> followed by T<sub>4</sub> at the tune of (86.67 %) which was statistically at par with T<sub>3</sub> having germination percentage (83.33 %) while the lowest germination percentage (63.33 %) was recorded in control. Maximum number of days for initial emergence (33.33) was observed in T<sub>6</sub> (control), followed by T<sub>1</sub> with (29.37) days to initial emergence while minimum number of days to initial emergence (26.86) were recorded in T<sub>5</sub>.

### Wingspan

The result in (Table 1) clearly shown that the concentration of GA<sub>3</sub> had significant effect on wingspan of new seedlings of sour orange. The wingspan (4.90 cm) was recorded in T<sub>5</sub> which was significantly higher than rest of the treatments, followed by T<sub>4</sub> (4.33 cm) which was statistically at par with rest of treatments rather than control and the minimum wingspan (3.43 cm) was recorded in control.

### Stem diameter

The data in (Table 1) revealed that the different concentration of GA<sub>3</sub> had significant effect on the stem diameter of sour orange. The stem diameter (1.77 mm) was obtained in T<sub>5</sub> which was significantly superior than the rest of the treatments, followed by T<sub>4</sub> with stem diameter (1.53 mm) which was statistically similar with T<sub>3</sub> having diameter (1.51 mm). While the lowest stem diameter (1.32 mm) was obtained from untreated seed (control) which was statistically on par with the T<sub>1</sub> and T<sub>2</sub> (1.33 mm, 1.45 mm) respectively.

**Table 1.** Impact of different GA<sub>3</sub> concentrations and scarification on germination percentage, days taken for initial germination, wingspan, and diameter on sour orange rootstock

Treatments	Germination (%)	Initial germination	Wingspan (cm)	Diameter (mm)
T <sub>1</sub>	73.33	29.37	4.00	1.33
T <sub>2</sub>	76.67	29.03	4.28	1.45
T <sub>3</sub>	83.33	28.35	4.27	1.51
T <sub>4</sub>	86.67	28.08	4.33	1.53
T <sub>5</sub>	96.67	26.86	4.90	1.77
T <sub>6</sub>	63.33	33.33	3.43	1.32
<b>SEm±</b>	<b>1.80</b>	<b>0.12</b>	<b>0.08</b>	<b>0.03</b>
<b>C.D. at 5%</b>	<b>9.61</b>	<b>0.64</b>	<b>0.44</b>	<b>0.14</b>

### Seedling Height

The data presented in (Table 2) revealed that, the application of GA<sub>3</sub> increased seedling height significantly. The results showed that the tallest plant height (4.99 cm) was observed in T<sub>5</sub>, which was statistically at par with T<sub>3</sub> and T<sub>4</sub> (4.9 cm, 4.57 cm) respectively. The shortest seedling height (3.88 cm) was found in T<sub>6</sub> (control) which was statistically the same as the remaining treatments.

### Root Number

Data recorded in (Table 2) indicated that different concentrations of GA<sub>3</sub> have influenced root numbers. The maximum root number (7.07) was observed in T<sub>5</sub> which was statistically at par with T<sub>4</sub> (6.23), followed by T<sub>3</sub>, T<sub>2</sub> and T<sub>1</sub> (5.97, 5.83 and 5.67) respectively. While the lowest root number (4.23) was obtained in untreated seed (control).

### Root Length

The results related to root length were shown in (Table 2) indicating that A<sub>3</sub> concentrations highly promoted root length. The highest root length (8.00 cm) was observed in T<sub>5</sub>, followed by T<sub>4</sub> with root length (6.20 cm) which was statistically at par with T<sub>3</sub>, T<sub>2</sub>, and T<sub>1</sub> (6.13 cm, 5.73 cm, and 5.50 cm) respectively. While the lowest root length (4.67 cm) was recorded in control, which was statistically on par with T<sub>1</sub>.

**Table 2.** Impact of different GA<sub>3</sub> concentrations and scarification on seedling growth, root number, and root length of sour orange rootstock

Treatments	Seedling height(cm)	Root number	Root length(cm)
T <sub>1</sub>	4.31	5.67	5.50
T <sub>2</sub>	4.23	5.83	5.73
T <sub>3</sub>	4.90	5.97	6.13
T <sub>4</sub>	4.57	6.23	6.20
T <sub>5</sub>	4.99	7.07	8.00
T <sub>6</sub>	3.88	4.23	4.67
<b>SEm±</b>	<b>0.09</b>	<b>0.18</b>	<b>0.18</b>
<b>C.D. at 5%</b>	<b>0.49</b>	<b>0.95</b>	<b>0.98</b>

## DISCUSSIONS

The attentive study of data from the (Table 1) displayed that different GA<sub>3</sub> concentrations and scarification significantly improved germination (%), initial germination emergence, wingspan (cm) and diameter (mm) in T<sub>5</sub> over the control. Gibberellic acid is one of the eco-friendly plant growth regulator on the embryo, leading to synthesis of hydrolytic enzymes e.i. amylase that hydrolyze food for embryo growth and enhance their germination (Leite *et al.*, 2003). The results are in agreement with the previous findings of (Kalalbandi *et al.*, 2003; Pawar *et al.*, 2010) The accelerated and enhanced germination in sour orange under different GA<sub>3</sub> concentrations and scarification may be due to increase in assimilation of water and increase in enzymes activity. (Meena *et al.*, 2003) also reported similar results and found that treated seed with the GA<sub>3</sub> @ 100 ppm encouraged early and highest germination. Another study reported by (Çalışkan *et al.*, 2012) and (Patel *et al.*, 2016) stated that the seed were subjected to 500 and 1000 ppm concentration of GA<sub>3</sub> achieved the maximum (100%) germination percentage and significantly decreased the time for initial emergence. Our results are in close conformity with the earlier researchers who reported that the application of GA<sub>3</sub> increased the seed germination and stimulated the seedling growth (Mtambalika *et al.*, 2014). Our findings are similar with the earlier study conducted by (Kumari *et al.*, 2007) who reported that the maximum germination percentage of

aoonla seeds (75.50 %) was recorded when seeds were treated with GA<sub>3</sub> 500 ppm over other treatments under Haryana conditions.

Our findings are also similar with reported literature of the fruit crops like Rangpur lime GA<sub>3</sub> application promoted and enhanced the morphological parameters including leaves number and stem diameter (Dala *et al.*, 2002). As well as (Qadri *et al.*, 2021) also reported that application of GA<sub>3</sub> @ 400 ppm increased growth parameters in rough lemon.

It is evident from the (Table 1) that GA<sub>3</sub> concentrations have significantly affected the seedling height, which may be due to the gibberellins that stimulate the plant's physiological activities and stimulate the rapid formation of new cells that are required for the better growth of seedlings (Sharma, 2012 & Sharma, 2016) therefore increase in plant height of seedling might be due to the increased in meristems tissues by the GA<sub>3</sub> applications. This investigation is confirmed by other researchers who stated that the shoot length, root length, fresh and dry weights of guava CV. SR-4 seedlings were highest when treated with 250 ppm GA<sub>3</sub> solution (Jholgiker *et al.*, 2017). Our results are similar to the findings of (Qadri *et al.*, 2021), who observed the maximum stem diameter (4.6 mm) with the application of GA<sub>3</sub> @ 400 ppm and matching findings reported by another researcher who stated that the application of GA<sub>3</sub> promoted vegetative growth (Hoda *et al.*, 2010).

The result reported in (Table. 2) indicated that maximum root number and root length were observed in T<sub>5</sub>, which was superior to other treatments, and the minimum was observed in control. It might be due the application of GA<sub>3</sub>, which caused vigorous root growth, hence, resulted in more roots per seedling. The GA<sub>3</sub> also accelerates the assimilation and translocation of auxins, which imparts better root growth, and vegetative characteristics of the plant as reported by (Pandey *et al.*, 2011). These results are in close agreement with (Patel *et al.*, 2017) in mango and also these results closely match with (Dilip *et al.*, 2017) in Rangpur lime; (Jain *et al.*, 2017) in Custard apple; (Parab *et al.*, 2017) in Papaya; Anjanawe *et al.*, 2013) in Papaya.

The increase in the length of roots may be due to the early formation of roots by GA<sub>3</sub>. Our results are in harmony with other researchers who stated that the maximum root length (8.82 cm) was obtained in T<sub>3</sub> @ 400 ppm GA<sub>3</sub> concentration. Similar results are also reported by (Dilip *et al.*, 2017) in Rangpur lime; (Singh *et al.*, 2017) in Kagzi lime; (Patil *et al.*, 2012) in Rangpur lime and they also reported regarding the influence of GA<sub>3</sub>, (Brijwal *et al.*, 2013; Shah *et al.*, 2013; Jholgiker *et al.*, 2017).

## CONCLUSION

It is known from the results of the present investigation that application of 250 ppm GA<sub>3</sub> with the mechanical scarification of sour orange seeds observed to be the best treatment for enhancing the growth and root parameters as well as rapid seedling establishment for early budding or grafting. However, using the intact seed should be avoided, because it reduces water absorption for proper imbibition and release some acids that inhibit germination. Availability of new technologies for plant improvement will make many plant hormone options available in the future. It is expected that growers will produce more seedling in short duration with access of plant hormone for citrus seeds. Highest benefit cost ratio was also observed in (1:6.124). Future scope the seed treated with GA<sub>3</sub> 250ppm identified in this investigation can be recommended commercially for overall growth of seedlings and Cost benefit ratio for sour orange rootstock.

## ACKNOWLEDGMENT

The authors are thankful to NUFA for providing the seeds and laboratory for the conducting of experiment and from all the professors of the agriculture faculty who helped me in providing proper guidance and relevant literatures for the present experiment.

## REFERENCES

- Anjanawe, S. R., Kanpure, R. N., Kachouli, B. K., & Mandloi, D. S. (2013). Effect of plant growth regulators and growth media on seed germination and growth vigour of papaya. *Annals of plant and soil research*, 15(1), 31-34.
- Brijwal, M., Kumar, R., & Mishra, D. S. (2013). Effect of pre-sowing treatments on seed germination of guava (*Psidium guajava* L.) under Tarai region of Uttarakhand. *Progressive Horticulture*, 45(1), 63-68.
- Çalışkan, O., Mavi, K., & Polat, A. (2012). Influences of presowing treatments on the germination and emergence of fig seeds (*Ficus carica* L.). *Acta Scientiarum. Agronomy*, 34, 293-297.
- Castle, W.S. (1982). Commercial citrus rootstocks in the United State.
- Choudhary, R. C., Kanwar, J., & Singh, P. (2022). Effect of Gibberellic acid (GA<sub>3</sub>) and growing media on seedling growth parameters of papaya (*Carica papaya* L.) cv. Pusa Nanha. *The Pharma Innovation Journal*, 11(1), 247-251.

- Dalal, S. R., Patil, S. R., Gonge, V. S., & Athawale, R. B. (2002). Effect of GA<sub>3</sub> and urea on growth of Rangpur lime seedlings in nursery.
- Dilip, W. S., Singh, D., Moharana, D., Rout, S., & Patra, S. S. (2017). Effect of gibberellic acid (GA) different concentrations at different time intervals on seed germination and seedling growth of Rangpur Lime. *J Agroeco Nat Resource Management*, 4, 157-165.
- Hoda, M. M., Abd EL-Rahman, G. F., & Abd el-rehman, M. E. (2010). Impact of gibberellic acid enhancing treatments on shortening time to budding of citrus nursery stocks. *Journal of American Science*, 6(12), 410-422.
- Jain, S., Sharma, T. R., Lal, N., Rangare, N. R., & Kumar, B. (2017). Effect of GA<sub>3</sub> and growing media on seed germination and growth of Custard apple (*Annonas quamosa* L.). *International Journal of Chemical Studies*, 5(4), 699-707.
- Jhologiker, P., Bade, M., & Sabard, A. (2017). Germination studies in different guava (*Psidium guajava* L.) cultivars. *International Journal of Current Microbiology and Applied Sciences*, 6(5), 2826-2829.
- Kalalbandi, B. M., Dabhade, R. S., Ghadge, P. M., & Vaishali Bhagat, V. B. (2003). Effect of gibberellic acid, naphthalene acetic acid and potassium nitrate on germination and growth of kagzi lime.
- Khatana, K. J., Jadav, R. G., & Nehete, D. S. (2010). *Influence Of Ga<sub>3</sub> On Germination And Growth Of Acidlime Cv. Kagzilime Seed (Citrus Aurantifolia Swingle) Under Field As Well As Net House Conditions* (Doctoral Dissertation, Anand Agricultural University).
- Leite, V. M., Rosolem, C. A., & Rodrigues, J. D. (2003). Gibberellin and cytokinin effects on soybean growth. *Scientia Agricola*, 60, 537-541.
- Meena, R. R., Jain, M. C., & Mukherjee, S. (2003). Effect of pre-sowing dip seed treatment with gibberellic acid on germination and purvivability of Papaya. *Annals of Plant Soil Research*, 5, 120-121.
- Mtambalika, K., Munthali, C., Gondwe, D., & Missanjo, E. (2014). Effect of seed size of *Azizia quanzensis* on germination and seedling growth. *International Journal of forestry Research*, 2014.
- Ollitrault, P., & Navarro, L. (2012). Citrus. *Fruit breeding*, 623-662.
- Parvin, P., Khezri, M., Tavasolian, I., & Hosseini, H. (2015). The effect of gibberellic acid and chilling stratification on seed germination of eastern black walnut (*Juglans nigra* L.). *Journal of nuts*, 6(01), 67-76.
- Patel, R. J., Ahlawat, T. R., Patel, A. I., Amarcholi, J. J., Patel, B. B. and Sharma, K. (2017). Growth of mango (*Mangifera indica* L.) rootstocks as influenced by presowing treatments. *Journal of Applied and Natural Science*, 9(1), 582-586
- Patel, R. J., Ahlawat, T. R., Singh, A., Momin, S. K., & Gavri, C. (2016). Effect of pre-sowing treatments on stone germination and shoot growth of mango (*Mangifera indica* L.) seedlings. *Int. J. Agric. Sci*, 8(52), 2437-2440.
- Patil, S. R., Sonkamble, A. M., & Khobragade, H. M. (2012). Influence of some growth regulators on germination and growth of Rangpur lime (*Citrus limonia* O.) seeds under shade net conditions. *Green Farming*, 3(6), 690-693.
- Qadri, R., Hussain, S., Akram, M. T., Khan, M. A., Khan, M. M., Hussain, K., ... & Khan, U. A. (2021). Impact of Different Growing Media and Gibberellic Acid (GA<sub>3</sub>) Concentrations on Rough Lemon (*Citrus Jambhiri*) Seed Germination and Its Growth Attributes. *International Journal of Modern Agriculture*, 10(2), 4471-4482.
- Rashmi Kumari, R. K., Sindhu, S. S., Sehrawat, S. K., & Dudi, O. P. (2007). Germination studies in aonla (*Emblica officinalis* Gaertn).
- Ratha, K., Rajwant, P., Kalia, K., Tewari, J. C., & Roy, M. M. (2014). Plant Nursery management: Principles and practices. *Jodhpur: Central Arid Zone Research Institute*, 40.
- Shah, R. A., Sharma, A., Wali, V. K., Jasrotia, A., & Plathia, M. (2013). Effect of seed priming on peach, plum and apricot germination and subsequent seedling growth. *Indian Journal of Horticulture*, 70(4), 591-594.
- Sharaf, M. M., Atawia, A. R., Bakry, K. A., & El-Rouby, M. Z. (2016). Effect of pre-sowing seeds soak in different GA<sub>3</sub> and ZnSO<sub>4</sub> solutions on germination and growth of Cleopatra mandarin and Rangpur lime rootstocks. *Middle East J Agric Res*, 5, 233-238.
- Sharma, D. K. (2016). Effect of plant growth regulators and scarification on germination and seedling growth of Chironji (*Buchanania lanzan* Spreng.). *Advances in Life Sciences*, 5(8), 3237-3241.
- Sharma, R. (2012) Effect of salicylic acid and gibberellic acid on seed germination and growth of pea. *International Journal of Plant Sciences (Muzaffarnagar)*, 7(2), 322-324
- Singh, R., Gurjar, B., & Baghel, S. S. (2017). Seed Germination and Seedling Vigour of Kagzi Lime (*Citrus aurantifolia* Swingle) As Influenced by Growth Regulators and Fungicide. *International Journal of Pure & Applied Bioscience*, 5(4), 2105-2109.
- Stejskalová, J., Kupka, I., & Miltner, S. (2015). Effect of gibberellic acid on germination capacity and emergence rate of sycamore maple (*Acer pseudoplatanus* L.) seeds.
- Vieira, A.R., M. D. G. G. C., Fraga, A.C., Oliveira, J. A., & Santos, C. D. D. (2002). Action of gibberellic acid (Ga<sub>3</sub>) on dormancy and activity of alpha – amylase in rice seed. *Revista Brasilia de Sementes*, 24,43-48
- Yadav, R. K., Prakash, O., Srivastava, A. K., Dwivedi, S. V., & Gangwar, V. (2022). Effect of plant growth regulators and thiourea on seed germination and seedling growth of Jatti Khatti (*Citrus jambhiri* Lush.). *The Pharma. Innovation J*, 11(6), 1393-1399.
- Zanotti, R. F., Dias, D. C. F. D. S., Barros, R. S., Silva, L. J. D., & Sekita, M. C. (2014). Germination of "Solo" papaya seeds treated with plant hormones. *Journal of Seed Science*, 36, 94-99.